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## Note

### $\alpha$ -Cyclodextrin as selective agent for the separation of *o*-, *m*- and *p*-nitrobenzoic acids by reversed-phase high-performance liquid chromatography

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Conventional high-performance liquid chromatographic (HPLC) and thin-layer chromatographic systems show little selectivity towards *m*- and *p*-nitrobenzoic acid (NBA), which have similar dissociation constants. The selectivity factor,  $\alpha_{m/p}$ , usually does not exceed *ca.* 1.2. The recently reported resolution of *o*-, *m*- and *p*-NBA in clathrate-containing systems<sup>1</sup>, although very selective ( $\alpha_{m/p} = 14-16$ ), has some major drawbacks, mainly that non-dissociated molecules of NBA but not anionic NBA<sup>-</sup> species are absorbed by the lipophilic channel structure of the  $\beta$ -Ni(NCS)<sub>2</sub>(4-methylpyridine)<sub>4</sub> clathrate sorbent. Acidic mobile phases have to be used, which results in a significant decrease in the clathrate column stability.

Our recent successful use<sup>2</sup> of  $\beta$ -cyclodextrin ( $\beta$ -CD) as a component for suitably modifying the separation properties of reversed-phase (RP) LC chromatographic systems and the known<sup>3</sup> differences between shifts of apparent dissociation constants of NBA isomers in the presence of  $\alpha$ -cyclodextrin ( $\alpha$ -CD) prompted us to try  $\alpha$ -CD to enhance the efficient separation of isomers of NBA.

## EXPERIMENTAL

### Reagents

$\alpha$ -Cyclodextrin was supplied by Chinoin (Budapest, Hungary). All other materials were of analytical or reagent grade and were used without further purification.

### Apparatus and procedure

Chromatographic measurements were performed using a Type 302 HPLC apparatus (Institute of Physical Chemistry, Polish Academy of Sciences, Warsaw, Poland) equipped with a 5- $\mu$ l high-pressure injection valve and a spectrophotometric detector (254 nm) with a Z-shaped passage (volume 8  $\mu$ l).

For HPLC use was made of stainless-steel columns (250  $\times$  4 mm I.D. and 100  $\times$  4 mm I.D.) slurry packed at 435 kg/cm<sup>2</sup> using the "balanced density" technique with 10  $\mu$ m LiChrosorb RP-18 (E. Merck, Darmstadt, G.F.R.).

pH measurements were performed using a Type N517 pH meter (Mera-Tronic, Poland).

The mobile phases were aqueous solutions of pH  $\approx$  3, 4 and 6, containing  $\alpha$ -CD at concentrations of *ca.*  $1 \cdot 10^{-3}$ ,  $2 \cdot 10^{-3}$ ,  $4 \cdot 10^{-3}$  and  $8 \cdot 10^{-3}$  M. Buffer solutions

were prepared by addition of 2.0 ml of concentrated orthophosphoric acid (85%) to 250 ml of water followed by titration to a given pH (checked with the pH meter) with 4 M sodium hydroxide solution.

Solutions of *o*-, *m*- and *p*-NBA dissolved in methanol ( $4 \cdot 10^{-3}$ – $5 \cdot 10^{-3}$  M) were injected onto the column. The flow-rate was 20  $\mu$ l/sec.

All chromatographic experiments were performed at  $20 \pm 1^\circ\text{C}$ .

## RESULTS AND DISCUSSION

Table I gives the measured capacity factors ( $k'$ ) of *o*-, *m*- and *p*-NBA and the observed selectivity factors,  $\alpha_{m/p}$  and  $\alpha_{p/o}$ , at different  $\alpha$ -CD concentrations and different pH values of the mobile phase. It can be seen that in the absence of  $\alpha$ -CD,  $\alpha_{m/p}$  does not exceed 1.16 (at pH 4.18); the highest value of  $\alpha_{m/p}$  (1.74) is observed at low pH (3.04) and high  $\alpha$ -CD concentration (0.0078 M); at high pH of the mobile phase the observed selectivity is low in both the presence ( $\alpha_{m/p} = 0.94$ ) and absence ( $\alpha_{m/p} = 1.04$ ) of  $\alpha$ -CD in the solution; and at intermediate pH, approximately within the limits  $pK_a - 2 < \text{pH} < pK_a + 2$ , high  $\alpha_{m/p}$  values may be attained by an appropriate admixture of  $\alpha$ -CD.

Fig. 1 shows a chromatogram of *o*-, *m*- and *p*-NBA obtained at a pH and  $\alpha$ -CD concentration considered to be optimal.

When studying chromatographic equilibria in the system (ion exchanger)<sub>solid</sub>–(sorbate + CD)<sub>dissolved</sub>, Uekama *et al.*<sup>4</sup> derived an equation that relates observed

TABLE I

VALUES OF CAPACITY FACTORS ( $k'$ ) OF *o*-, *m*- AND *p*-NBA AND SELECTIVITY FACTORS,  $\alpha_{m/p}$ ,  $\alpha_{p/o}$ , OBTAINED FROM CHROMATOGRAPHIC MEASUREMENTS AT VARIOUS pH VALUES AND CD CONCENTRATIONS

| $\alpha$ -CD<br>concentration<br>(mmole/l) | pH   | $k'$ of NBA |            |            | $\alpha_{m/p}$ | $\alpha_{p/o}$ |
|--|------|-------------|------------|------------|----------------|----------------|
|  |      | <i>o</i> -  | <i>m</i> - | <i>p</i> - |                |                |
| 0.0  | 2.95 | 4.99        | 33.94      | 32.38      | 1.05           | 6.49           |
| 0.0  | 4.18 | 1.63        | 8.60       | 7.39       | 1.16           | 4.53           |
| 0.0  | 5.92 | 1.28        | 3.89       | 3.50       | 1.11           | 2.73           |
| 0.9913                                     | 3.00 | 3.66        | 23.52      | 18.68      | 1.26           | 5.10           |
| 0.9892                                     | 3.87 | 1.69        | 9.61       | 8.29       | 1.16           | 4.90           |
| 0.9845                                     | 5.95 | 1.25        | 3.53       | 3.18       | 1.11           | 2.54           |
| 1.9509                                     | 3.05 | 3.58        | 19.16      | 15.27      | 1.25           | 4.26           |
| 1.9451                                     | 4.25 | 1.41        | 6.58       | 5.60       | 1.17           | 3.97           |
| 1.9350                                     | 6.05 | 1.27        | 3.42       | 3.30       | 1.04           | 2.60           |
| 3.9555                                     | 3.14 | 2.84        | 15.03      | 10.02      | 1.50           | 3.53           |
| 3.9449                                     | 4.57 | 1.26        | 4.25       | 3.79       | 1.12           | 3.01           |
| 3.9268                                     | 5.92 | 1.16        | 3.16       | 3.01       | 1.05           | 2.59           |
| 7.8745                                     | 3.04 | 3.11        | 12.01      | 6.89       | 1.74           | 2.21           |
| 7.8545                                     | 3.87 | 1.42        | 5.88       | 4.33       | 1.36           | 3.05           |
| 7.8208                                     | 6.00 | 1.09        | 2.60       | 2.77       | 0.94           | 2.54           |

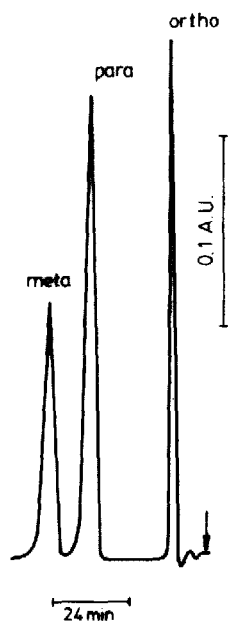


Fig. 1. Elution curve of a mixture of  $5 \cdot 10^{-3} M$  *o*-, *m*- and *p*-NBA with an aqueous mobile phase of pH 2.93 and  $[α\text{-CD}] = 0.0099 M$ . Column:  $250 \times 4$  mm I.D. LiChrosorb RP-18 ( $10 \mu\text{m}$ ).

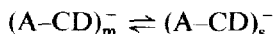
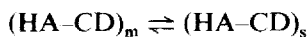
retention times ( $t_{\text{obs}}$ ) of ionic species and the concentration of cyclodextrin in the mobile phase ( $[\text{CD}]$ ):

$$t_{\text{obs}} = \frac{t'_0 + t_c K_c [\text{CD}]_m}{1 + K_c [\text{CD}]_m}$$

where  $t'_0$  is the retention time of the sorbate,  $t_c$  that of the sorbate-CD complex and  $K_c$  is the stability constant of the 1:1 complex. If acid-base equilibria of the sorbate in the mobile phase:



and also adsorption of both neutral and anionic species on a reversed phase:



where the subscripts *s* and *m* denote the stationary and mobile phase, respectively, are taken into account then the following expression can be derived for the equilibrium distribution constant of HA:

$$C = \frac{[\text{HA}]_s + [\text{A}^-]_s + [\text{HA-CD}]_s + [(\text{A-CD}^-)]_s}{[\text{HA}]_m + [\text{A}^-]_m + [\text{HA-CD}]_m + [(\text{A-CD}^-)]_m}$$

and for the retention time:

$$t_{\text{obs}} = \frac{t_{\text{HA}} + t_{\text{A}^-} \frac{K_a/[\text{H}^+]}{1 + K_a/[\text{H}^+]} + t_{\text{HA-CD}} \frac{K^\circ [\text{CD}]}{1 + K_a/[\text{H}^+]} + t_{(\text{A-CD})^-} \frac{K^- [\text{CD}] K_a/[\text{H}^+]}{1 + K_a/[\text{H}^+]} + K^- [\text{CD}] K_a/[\text{H}^+}}{1 + K_a/[\text{H}^+]} \quad (1)$$

where  $t$  denotes retention time and the subscripts obs, HA, A<sup>-</sup>, HA-CD and (A-CD)<sup>-</sup> refer to overall (measured) values and to the retention of neutral, anionic, neutral-complexed and anionic-complexed molecular species, respectively, and  $K$  is the acidity constant ( $K_a$ ) or stability constant of CD (1:1) complexes of neutral

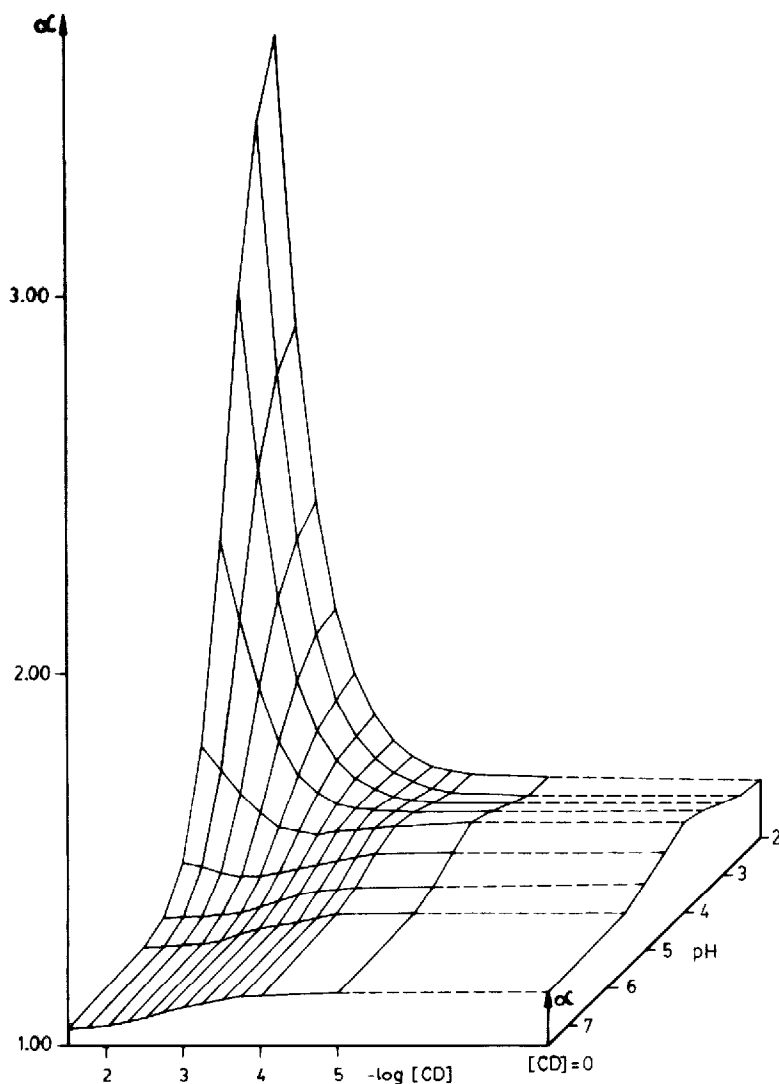


Fig. 2. Selectivity factor,  $\alpha_{mip}$ , as a function of pH and  $\log[\alpha\text{-CD}]$  calculated from eqn. 1 and the data in Table II.

$$\left( K^{\circ} = \frac{[\text{HA-CD}]_m}{[\text{HA}]_m[\text{CD}]_m} \right)$$

and anionic

$$\left( K^{-} = \frac{[(\text{A-CD})^{-}]_m}{[\text{A}]_m[\text{CD}]_m} \right)$$

species.

For numerical procedures it is useful to expand the non-linear function  $t_{\text{obs}} = t([\text{H}^+][\text{CD}])$  into a Taylor series and neglect the non-linear components. In this way we calculated the parameters  $K^{\circ}$ ,  $K^{-}$  and individual capacity factors  $k'_{\text{HA}}$ ,  $k'_{\text{A}^{-}}$ ,  $k'_{\text{HA-}\alpha\text{-CD}}$  and  $k'_{(\text{A-}\alpha\text{-CD})^{-}}$  corresponding to adsorption on a reversed phase of individual species by using a set of twelve values for  $t_{\text{obs}}$  measured at different pH values and  $[\alpha\text{-CD}]$  concentrations for each isomer. Some results are given in Table II.

TABLE II

CALCULATED STABILITY CONSTANTS ( $K^{\circ}$ ,  $K^{-}$ ) AND CAPACITY FACTORS [ $k'_{\text{HA}}$ ,  $k'_{\text{A}^{-}}$ ,  $k'_{\text{HA-}\alpha\text{-CD}}$ ,  $k'_{(\text{A-}\alpha\text{-CD})^{-}}$ ] FOR  $\alpha\text{-CD}$  COMPLEXES OF *m*- AND *p*-NBA

| Compound      | $k'_{\text{HA}}$ | $k'_{\text{A}^{-}}$ | $k'_{\text{HA-}\alpha\text{-CD}}$ | $k'_{(\text{A-}\alpha\text{-CD})^{-}}$ | Literature $pK_a$ values <sup>5</sup> | $K^{\circ}$   | $K^{-}$                              |
|---------------|------------------|---------------------|-----------------------------------|--|---------------------------------------|---|--------------------------------------|
| <i>m</i> -NBA | 42.85 ± 0.10     | 3.63 ± 0.02         | 5.62 ± 0.34                       | 2.98 ± 0.05                            | 3.47                                  | 408 ± 21<br>(lit.: 155 <sup>6</sup> ;<br>152 <sup>7</sup> ) | 486 ± 16<br>(lit.: 50 <sup>7</sup> ) |
| <i>p</i> -NBA | 37.30 ± 0.28     | 3.19 ± 0.04         | 0.28 ± 0.27                       | 2.86 ± 0.04                            | 3.41                                  | 473 ± 16<br>(lit.: 155 <sup>6</sup> ;<br>490 <sup>7</sup> ) | 359 ± 16<br>(lit.: 75 <sup>7</sup> ) |

These results seem interesting but their reliability can hardly be evaluated in view of large discrepancies in literature data on the association constants of  $\alpha\text{-CD}$  inclusion complexes of NBA and even on dissociation constants. Nevertheless, some qualitative conclusions may perhaps be given as reasonable. In particular, the decrease in adsorption on an RP-18 stationary phase of  $\alpha\text{-CD}$  inclusion complexes of NBA molecules, if related to the adsorption of NBA isomers themselves ( $k'_{\text{HA-CD}}$  compared to  $k'_{\text{HA}}$ ), is very significant:  $k'_{\text{HA-CD}}$  found for the *o*-NBA is 35% of  $k'_{\text{HA}}$ , for *m*-NBA only 13% of  $k'_{\text{HA}}$  and for the *p*-isomer  $k'_{\text{HA-CD}}$  is zero within the limits of error.

The sequence of the calculated capacity factors for individual species is as follows:  $k'_{\text{HA}} > k'_{\text{A}^{-}}$ ;  $k'_{\text{HA}} > k'_{\text{HA-}\alpha\text{-CD}}$ ; and  $k'_{\text{A}^{-}} > k'_{(\text{A-}\alpha\text{-CD})^{-}}$ .

A deeper physico-chemical interpretation of these results requires further studies, which are in progress.

Eqn. 1 with the derived values of the parameters listed above can be used to illustrate the relationship between  $\alpha_{m/p}$  and the concentration of  $\alpha\text{-CD}$  and the pH of the mobile phase. The result is shown in Fig. 2.

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## REFERENCES

- 1 W. Kemula, D. Sybilska and J. Lipkowski, *J. Chromatogr.*, 218 (1981) 465.
- 2 J. Dębowski, D. Sybilska and J. Jurczak, *J. Chromatogr.*, 237 (1982) 303.
- 3 A. Connors and J. M. Lipari, *J. Pharm. Sci.*, 65 (1976) 379.
- 4 K. Uekama, F. Hirayama, S. Nasu, N. Matsuo and T. Irie, *Chem. Pharm. Bull.*, 26 (1978) 3477.
- 5 F. J. Dippy and R. H. Lewis, *J. Chem. Soc.*, (1936) 644.
- 6 B. Casu and L. Rava, *Ric. Sci.*, 36 (1966) 733.
- 7 R. I. Gelb, L. M. Schwartz, B. Cardelino, H. S. Fuhrman, R. F. Johnson and D. A. Laufer, *J. Amer. Chem. Soc.*, 103 (1981) 1750.